



Optimizing of the quality of rose grown with varying ratios and periods of Red: Blue light-emitting diodes in commercial greenhouse

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ABSTRACT

Purpose: This research investigated the effect of different regimes of supplementary LED light on physiological and morphological traits of two cultivars of cut Roses. **Research method:** In this study, treatments included cultivars (Allstar and Dolcevit) and nine light regimes including (Control, LED night from 5 pm to 7 am, LED day from 7 am to 5 pm, LED night day (24 hours light) and LED dark (24 hours, without sunlight) which were all applied in two intensities of 2000 and 4000 lux. **Findings:** The results showed that the LED light regime had a positive effect on morphological traits such as the number of buds and mean harvest, length, diameter and fresh weight of flowers. LED day 4000 lux improved bud diameter and mean harvest by 18% and 112% respectively compared with control in Dolcevit cultivar. The light regime also caused a significant improvement in physiological characteristics so that in Allstar cultivar LED day 4000 lux, chlorophyll a, b, total, and carotenoid were increased by 66%, 60%, 63%, and 64% respectively compared with control. The vase life in Allstar cultivar by LED night day 4000 lux intensity and Dolcevit cultivar by LED day 4000 lux were 44.5% and 133.2% higher than the control treatment, respectively. **Research limitations:** There was no limitation. **Originality/Value:** The results showed that LED night day with 4000 lux intensity had the best results in vase life in Allstar cultivar and LED day supplementary light with 4000 lux intensity increased the quantity and quality characteristics of roses.

INTRODUCTION

Rosa hybrida is one of the most popular cut flowers. Several elements influence the quality and postharvest life of roses after they have been cut (Shi et al., 2021). These factors are classified into two categories: preharvest and postharvest. The most important preharvest stage parameters that have a significant influence on appearance quality, physiological features, and vase life of flowers are irrigation conditions, mineral nutrients, and the amount and quality of light. Among these elements, light is crucial for floral quality, biosynthesis, and pigment content (Rasouli et al., 2015).

Livadariu et al. (2023) state that LED lighting has become more prevalent in the past few decades for the commercial breeding of various economically important species in horticulture and agriculture. LED lights are long-lasting, have high radiative efficiency, and switch quickly. Furthermore, they enable the selection and customization of output spectrum characteristics, which meet the demands of the plant and allow for high-quality harvests including antioxidant capacity (Carotti et al., 2021). In recent studies by Song et al. (2022), it has been reported that different light wavelengths have the ability to regulate various plant processes, including photosynthesis, germination, flowering, biomass accumulation, and phytochemical synthesis. Research indicates that red light plays a crucial role in the development of the photosynthetic apparatus and influences morphogenesis through light-induced transformations of the phytochrome system. On the other hand, blue light can impact chlorophyll biosynthesis, stomata opening, and photomorphogenesis (Song et al., 2022; Sakurako et al., 2021). As red and blue light are the primary absorption peaks of photosynthetic pigments in plant leaves, they have a significant impact on plant photosynthesis. However, it is important to note that a single red or blue light is insufficient for normal plant growth. Studies suggest that a specific proportion of red and blue light is necessary for optimal plant growth (Song et al., 2022).

The effect of light on the proportion of plant hormones is one of the processes in the effect of light on plant growth and morphology. Kurepin et al. (2010) discovered that long-term culture under changing light conditions has an effect on internal hormone levels, particularly auxin, and induces homeostasis. Furthermore, it appears that high-intensity blue light damages or does not synthesize auxin. According to Cioć et al. (2022), the application of red and red:blue light in gerbera flowers led to a decrease in tissue auxin levels. In contrast, the use of blue LED light resulted in a lowered the shoot multiplication rate and their height, but it induced the highest content of gibberellins in the final stage of the culture.

According to Alsanius et al. (2017), red LED light 80% + blue 20% with 16 hours of illumination and intensity of 7000 lux improved sunflower plant height and stem diameter. Schroeter-Zakrzewska and Pradita (2021) discovered that Chrysanthemum plants grown under red + blue light had the highest leaf greenness index (SPAD) value and the shortest cuttings with the longest roots. Additionally, white + blue light significantly affected most of the growth parameters, except for plant height and the number of leaves.

Cut flower post-harvest senescence is a dynamic process involving physiological and biochemical changes that are governed by a cell death program. Carnation senescence's physiological and biochemical features have already been characterized, and conditions during mother plant growth, storage and handling, environment, and phytohormones all play roles in senescence regulation (Aalifar et al., 2020). Postharvest or vase life as a commercial attribute impacts the market's flexibility at any given time, particularly for cut flowers. The limited vase life of cut rose flowers is related to physio-chemical processes that influence aging. Water loss and wilting during transit have a significant impact on these characteristics.

Water scarcity and the resulting premature senescence result in poor cut flower quality and market loss, according to numerous reports (Alaey et al., 2011).

Optimizing the quality of roses in commercial greenhouses through different ratios and durations of LED lighting is an important research topic in agriculture today. As it has a significant effect on the quality and performance of roses in greenhouses. By determining the optimal ratio and duration of red:blue LED lighting, growers can ensure that their roses receive the right amount and type of light, which can lead to improved quality, marketability, and vase life. This, in turn, can lead to a reduction in energy consumption and an increase in profitability for producers. In this regard, the present research investigates the influence of supplemental LED light on the physiological and morphological features of two cut rose cultivars, "Allstar" and "Dolcevida," in a commercial greenhouse under different LED light regimes. The study also examines the effect of pretreatment of roses with LED on their vase life. The findings of this research can provide valuable insights into the optimal lighting conditions for rose growth and inform future research and innovation in the field.

MATERIALS AND METHODS

Culture conditions and light treatments

This experiment was conducted in the Sivan Energy commercial rose greenhouse in Dahagan, Isfahan province, as a factorial experiment in the form of a completely randomized design (CRD). This study included three replications and focused on two rose cultivars, Allstar and Dolcevida. Allstar and Dolcevida rose plants were imported from the Dutch company De Ruiter, Amstelveen, and the Netherlands, whose standard morphological characteristics are shown in Table 1 according to the manufacturer's information. Then, the hydroponic bed was prepared and the rose bushes, which were approximately 25 cm in size and had 2-3 leaves, were planted in the perlite bed with a distance of 19 cm between the bushes. To start establishing the plants, they were watered for two weeks, and then NPK fertilizer (20-20-20) was given to the rose plants for one month and the plants were fed daily with the nutrient solution, the information of which is included in Table 2 (Nikbakht & Ashrafi, 2019)

Light regime treatments (red wavelength 60%+blue wavelength 40%) included 9 levels: Control (without using LED light), LED night (5 pm to 7 am) with two intensities of 2000 and 4000 lux+sunlight, LED day (7 am to 5 pm) with two intensities of 2000 and 4000 lux+sunlight, LED day and night (24 hours of light) with two intensities of 2000 and 4000 lux+sunlight and LED dark (24 hours without sunlight) with two intensities of 2000 and 4000 lux (Table 3).

Five months after the growth and full establishment of the plants, LED lamps were placed at a distance of approximately one meter from the substrate to provide the desired light intensity on the leaf surface. The light intensity of 2000 ($35\mu\text{molm}^{-2}\text{s}^{-1}$) and 4000 ($75\mu\text{molm}^{-2}\text{s}^{-1}$) lux was adjusted by adjusting the amount of use of these lamps in the panels, and the wavelength of the lamps used was 660 (red), and 460 nm (blue) (Fig. 1 and Fig. 2). The control treatment received only sunlight and no LED light was applied. In the LED night treatment, LED treatments applied during the day and night, the light intensity of the LED lamps was 2000 and 4000 lux along with the sunlight intensity. But in the LED dark treatment, the plant environment was darkened with black covers to minimize the effect of sunlight intensity, then the light regimes were applied on Allstar and Dolcevida rose cultivars with three repetitions, each repetition including six rose bushes. The light treatments were applied for about three months and during this period we had six to seven peak harvests, where the measured parameters were obtained from the last harvest.

Table 1. Specifications of Allstar and Dolcevita cultivars in standard conditions.



| Cultivar | Shape | Bud length (cm) | Stem length (cm) | Vase life (days) | Production in square meters per year (flower branch) |
|-----------|---|-----------------|------------------|------------------|--|
| Allstar |  | 5.4-5.5 | 40-60 | 10-12 | 180-220 |
| Dolcevita |  | 5-6 | 50-80 | 7-10 | 150-200 |

Table 2. Hydroponic nutrition solution formula in this experiment.

| (μ M) | Micro elements | (mM) | Macro elements |
|------------|----------------|------------|--------------------------------|
| 30-50 | Fe | 11/25-11/5 | NO ₃ |
| 5-6 | Mn | 1-1/5 | NH ₄ |
| 4-5 | Zn | 1/25-1/3 | H ₂ PO ₄ |
| 30-40 | B | 4/5-5 | K |
| 0/60-0/75 | Cu | 3/5-4 | Ca |
| 0/5-0/6 | Mo | 1/25-1/3 | Mg |
| 1/6 | EC | 1/25-1/3 | SO ₄ |
| 5/5 | pH | 1/25-1/3 | HCO ₃ |

Table 3. Introduction of experimental treatments.

| Cultivar | Light regimes |
|---|---|
| Allstar/Dolcevita | Control (Without LED light) |
| | LED night(5 pm to 7 am) with 2000 lux intensity + sunlight |
| | LED night(5 pm to 7 am) with 4000 lux intensity + sunlight |
| | LED day (7 am to 5 pm) with 2000 lux intensity + sunlight |
| | LED day (7 am to 5 pm) with 4000 lux intensity + sunlight |
| | LED night day (24 hours light) with 2000 lux intensity + sunlight |
| | LED night day (24 hours light) with 4000 lux intensity + sunlight |
| | LED Dark (24 hours, without sunlight) with 2000 lux intensity |
| LED Dark (24 hours, without sunlight) with 4000 lux intensity | |

LED include red wavelength 60% + blue wavelength 40%



Fig. 1. LED lamps installed in the greenhouse.

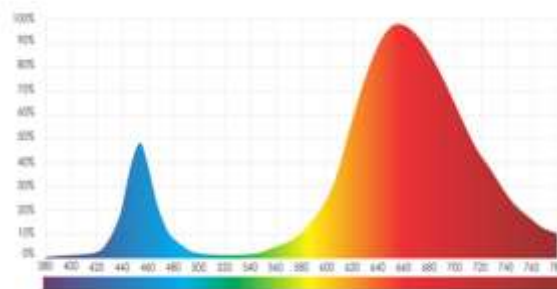


Fig. 2. Wavelengths used in LED boards.

Harvesting and measuring the growth parameter

Throughout the experimental stages from February 15 for 10 weeks in order to check the effect of the treatments, the number of flowers that were produced in each treatment from each plant was picked and recorded. The number of buds was counted one week after the treatment, regardless of whether the resulting buds changed into harvestable flowers, and the counts were averaged for each repetition individually and recorded weekly for ten weeks, with the results given for each replication after averaging. The length of the stem from under the sepal to the cut point (ten weeks) was also measured with a ruler and reported for each replication.

For each repeat of three flowers, the weight of cut flowers was measured in three phases at two-week intervals using a scale with an accuracy of 0.01 g for 30 cm long cut flowers, and its size was measured for each repetition and reported after averaging. After weighing three flowers for each replication, they were packaged in paper envelopes and placed in a 70-degree oven for 48 hours to determine the dry weight of cut flowers with a length of 30 cm. The number of leaves on each bloom was counted in each repeat and averaged for each repetition, and this experiment was repeated ten times with a one-week interval. The leaf area of the plants was measured using an Alborz Andisheh Technologies Co. leaf area measurement device, model Winarea-UT-11, in each replication of 8-10 leaves.

Physiological factors

A sample of 5 g of the tissue was homogenized with 80% acetone; the residue was filtered and adjusted to 10 mL; the absorbance reading was done at 476, 646, and 663 nm using the spectrophotometer (Model UV 160A- Shimadzu Corp., Kyoto, Japan) (Arnon, 1949). The content of carotenoids was calculated based on the formula (1) presented by Pérez -Grajales et al. (2019).

$$\text{Total chlorophyll} = \text{Chlorophyll a} + \text{Chlorophyll b}$$

$$\text{Carotenoids} = 100(A_{476}) - 3.27(\text{mg/g Chl. a}) - 104 (\text{mg/g Chl. b})/227 \quad (1)$$

The spectrometric method by Rapisarda et al. (2000) was used to determine anthocyanin. Fresh leaves (1.0 g) were crushed with 20 mL alcohol (60 %; pH=3.0) and heated the samples on hot water for 2 h; after cooling the samples certain volume of sample solution was used for reading at 535 nm anthocyanin content (mg/100 g FW).

The chlorophyll index of mature plant leaves was used by the chlorophyll meter (SPAD-502 plus, Japan). For this goal, three readings were carried out from each plant on three separate leaves (a total of 9 readings per replicate), and then the average was registered (Dezhabad & Haghghi, 2020). Maximum photochemical quenching F_v/F_m was measured by chlorophyll fluorescence (model OS-30, Minolta Corp). The portable fluorescence monitoring system (Hansatech Instruments LTD RS232) was used for chlorophyll fluorescence determination. The instrument clamp was placed on each leaf for 30 min to complete the dark adaptation. The initial (F_0), maximum (F_m), and maximum quantum efficiency of the photosystem-II (F_v/F_m) were reported according to Maxwell and Johnson (2000).

Vase life experiment

The appropriate flowers were selected early in the morning using sterilized scissors and immediately placed in water. They were then taken to Isfahan University of Technology and placed in a cold room at 4°C for 8 hours to absorb water and cool the flowers. The next morning, all flowers were cut at a distance of 30 cm from the end of the flower bud and immediately placed in 500 ml containers containing distilled water that had been disinfected

with 70% alcohol at a temperature of 24±2°C and a humidity of 40±5%, and were kept in 13 hours of light and 11 hours of darkness.

To determine carbohydrates content, the method of Hedg and Hofreiter (1962) was used. In this way, 100 mg of the sample was placed in a boiling bath with 5 mL of 2.5 N hydrochloric acid for 3 hours. Then, a few drops of 2% sodium carbonate were added to each sample. Add 25 mL of distilled water to the resulting solution to reach a volume of 50 ml. 0.1 mL of the resulting solution was removed and made up to 10 ml with distilled water. In the next step, 2 mL of anthrone reagent was added to the solution and placed in a boiling water bath for 1 minute. Then the resulting solution was cooled at the laboratory temperature and its light absorption was read at 630 nm by a spectrophotometer model (Shimadzu UV106A) (Hedge & Hofreiter, 1962). The amount of carbohydrates in the studied sample was estimated in terms of mg/g of dry sample weight using a standard curve. 0, 0.2, 0.4, 0.6, 0.8, and 1 ml of standard were used to prepare the standard curve.

The vase life of the flowers was considered from the time of harvesting to the bending of the neck or falling of the petals and was expressed as a number of days (Shi et al., 2021).

The speed of flower opening was measured by calipers on days 0, 2, 5, and 8, and the maximum amount of flower opening was subtracted from the diameter of the flower on day 0 and divided by the number of days, which was calculated in millimeters per day.

Flower water content was obtained by dividing the difference between the heavier and dry weight of the flower by its heavier weight. This experiment was performed twice in two weeks and for each repetition, the calculation and its numbers were reported separately.

Statistical analysis

The study applied following a factorial experiment in a complete randomized design (CRD) with three replications. The data underwent analysis using SAS program version 9.1 (SAS Institute, Cary, NC). The data performed a two-way analysis of variance (ANOVA), and the means were examined for statistical significance using the least significant difference (LSD) test at a significance level of P<0.05. Principal component analysis (PCA) was carried out using Statgraphics Centurion, Version XVI.

RESULTS AND DISCUSSION

The results of variance analysis are presented in Table 4. The main effects are not shown due to the significance of the interaction effect and the rising length of the article.

Table 4. Analysis of variance of main and interaction effects of treatments.

| Source | df | Bud number | Mean harvest weekly | Cut flower height | Bud length | Bud diameter | Number of leaves | Stem diameter | Flower quality |
|----------------|----|------------|---------------------|-------------------|------------|--------------|------------------|---------------|----------------|
| Cultivar | 1 | 0.4877 n.s | 0.09086 ** | 145.366 n.s | 5.59185 ** | 1.70951 ** | 37.9178 ** | 0.03393 ** | 0.42934 n.s |
| Light | 8 | 10.1285 ** | 0.20967 ** | 188.908 ** | 0.80121 ** | 0.34802 ** | 8.0516 ** | 0.3337 ** | 3.26838 ** |
| Cultivar×Light | 8 | 0.5771 ** | 0.05788 ** | 102.617 * | 0.03617 * | 0.0469 * | 3.2455 ** | 0.00503 ** | 0.60103 * |
| Error | 34 | 0.0889 | 0.01218 | 53.881 | 0.0362 | 0.02864 | 0.2841 | 0.0004 | 0.25421 |
| CV | | 6.2 | 10.4 | 13.52 | 3.68 | 4.56 | 5.43 | 2.23 | 6.51 |

ns: not significant, * significant at P ≤0.05 and ** significant at P ≤0.01 probability level, df: degree of freedom.

Table 4. (Continued).

| Source | df | Greenness | Chlorophyll fluorescence | Fresh weight of cut flowers | Dry weight of cut flowers | Leaf surface | Thrips pest | Chlorophyll a of leaves |
|----------------|----|------------|--------------------------|-----------------------------|---------------------------|---------------|-------------|-------------------------|
| Cultivar | 1 | 433.217 ** | 0.00089 n.s | 390.48 ** | 11.6111 ** | 2.325E+10 n.s | 0.0741 n.s | 0.00258 n.s |
| Light | 8 | 51.644 ** | 0.00263 ** | 299.717 ** | 10.1983 ** | 2.655E+10 * | 20.713 ** | 0.01543 ** |
| Cultivar×Light | 8 | 15.011 ** | 0.00074 * | 38.426 ** | 1.1937 ** | 2.700E+10 * | 0.4907 * | 0.01058 ** |
| Error | 34 | 3.769 | 0.0005 | 3.684 | 0.2004 | 2.64E+10 | 0.1991 | 0.00125 |
| CV | | 3.87 | 2.9 | 4.8 | 6.42 | 4.56E+02 | 9.41 | 11.57 |

ns: not significant, * significant at P ≤0.05 and ** significant at P ≤0.01 probability level, df: degree of freedom.

Table 4. (Continued).

| Source | df | Chlorophyll b of leaves | Total chlorophyll of leaves | Carotenoids of leaves | Anthocyanin of petal | Carbohydrate of leaves | Vase life | Speed of flower bud opening | Flower water content |
|----------------|----|----------------------------|-----------------------------------|--------------------------|-------------------------|---------------------------|------------|-----------------------------------|----------------------------|
| Cultivar | 1 | 0.00435 n.s | 0.02664 n.s | 0.00139 n.s | 708460 ** | 0.21426 ** | 13.0046 ** | 0.1057 n.s | 0.01051 n.s |
| Light | 8 | 0.0068 ** | 0.10137 ** | 0.00728 ** | 3398 * | 0.24455 ** | 9.3542 ** | 0.80612 ** | 1.19501 * |
| Cultivar×Light | 8 | 0.00282 * | 0.06262 ** | 0.0045 ** | 8603 ** | 0.02369 ** | 0.9421 * | 0.30697 * | 0.45356 * |
| Error | 34 | 0.00149 | 0.00994 | 0.00056 | 1603 | 0.00705 | 0.509 | 0.16335 | 0.89472 |
| CV | | 16.69 | 12.15 | 12.13 | 27.29 | 7.58 | 6.67 | 16.39 | 1.14 |

ns: not significant, * significant at $P \leq 0.05$ and ** significant at $P \leq 0.01$ probability level, df: degree of freedom.

The effect of LED on morphological factors of two rose cultivar

The number of buds, flower harvest (weekly), cut flower height, bud length, bud diameter

In the interaction effects of the light regime with the cultivar, the Allstar produced the most bud number in the LED day 2000 lux intensity treatment, which was 37% more than the control. Dolcevitaa produced the highest number of buds with LED night 4000 lux which was 42% more than the control (Table 5). A positive correlation was detected at the 1% level between bud number and bud length, diameter of bud, number of leaves, flower number, greenness, fresh weight, dry weight, carbohydrate, and vase life.

The results of study Park and Jeong (2020) indicated that a 4-h supplementation of blue light with a wavelength of 450 nm during the photoperiod increases flower bud formation in chrysanthemum. Light treatments were found to increase the amount of collected buds and flowers in this study. Heo et al. (2003) found that irradiating cyclamen (*Cyclamen persicum*) with red-blue light at a 1:1 ratio with a light intensity of 5500 lux for 12 hours can enhance the number of flower buds compared to the control and other monochromatic kinds, as well as make the blooms survive longer. On the facility for approximately 20 days longer than the control.

The interaction effect of light treatments with cultivar, the highest number of flowers harvested in Allstar was observed in LED dark 2000 lux treatment, which was 17% compared with the control. In the Dolcevitaa, the highest number of harvested flowers was observed in the treatment of LED day 4000 lux intensity, which increased by 105% compared to the control (Table 5). There is a positive correlation at the level of 1% with chlorophyll a, total chlorophyll, and relative water content at a 5% level, and a negative correlation was observed with thrips density index (was scored between lower 1 to higher 10 observation).

We can expect varied reactions in the formation of flowers in plants if we use different light spectrums and durations of light presence. The amount of internal auxin in plants can also be regulated by the quality of light. According to one study, the quality of light influences the activity of IAA oxidase and causes an increase in the length of roots and stems, which is most likely produced by a change in the amount of internal auxin in the plant (Iacona & Muleo, 2010). According to Hao et al. (2016), red-blue supplementary light at a ratio of 1:1 with a light intensity of 5000 lux on the cloud plant (*Houstonianum* cv. Blue Field Ageratum) increased the number of flowers. Gao et al. (2023) conducted an experiment which revealed a lower R:FR ratio of 54%:46% under natural light treatments compared to artificial light treatments. Interestingly, the high proportion of far-red light in natural light was found to promote the flowering number in *Crocus sativus* L. plants.

In the interaction effect of light treatment with cultivar, the highest cut flower height was observed in the Allstar cultivar in the treatment of LED dark 2000 Lux, which was 45% higher than the control. In the Dolcevitaa cultivar, the highest flower height harvested in LED night 2000 lux treatment was observed, which was a 40% increase in height compared with the control (Table 5). Thrips pest symptom and water content showed a positive and significant correlation at the 1% level, and with the weight, leaf area, and relative water content at the 5% level.

In the interaction effect of light treatment with cultivar, the maximum bud length was observed in the Allstar cultivar in LED night day with 4000 lux intensity, which was 20% more than the control. In the Dolcevita cultivar, the maximum bud length was observed in the LED night 4000 lux, which is a 13% increase in bud length compared with the control (Table 5). The bud length index with the indices of bud number, bud diameter, number of leaves, fresh weight, dry weight, and carbohydrate, a positive correlation was observed at the level of 1% and anthocyanin index at the level of 5%, a positive and significant correlation was observed between the traits.

In the interaction effect of light treatment with the cultivar, the largest bud diameter was observed in the Allstar cultivar in LED night 4000 lux treatment, which was 20% more than the control. In the Dolcevita cultivar, the maximum bud length was observed in LED night day 2000 lux, which is an 18% increase in bud diameter compared with the control (Table 5). The bud diameter with indicators of bud length, stem diameter, number of leaves, fresh weight, dry weight, and carbohydrates, a positive correlation was observed at the level of 1% and a positive and significant correlation was observed with the number of buds at the level of 5%.

Kurepin et al. (2010) discovered that long-term cultivation under different light conditions has an effect on internal hormone levels, particularly auxin, and causes homeostasis. Furthermore, it appears that high-intensity blue light destroys or does not produce auxin, altering the auxin-cytokinin ratio. Acetic acid is broken down, resulting in less longitudinal growth and less internode growth (Lee & Palsson, 1994). Alternating red and blue LED light that was on for 8 hours and off for 16 hours increased plant height in lettuce (*Lactuca sativa*) compared to illumination for 4, 2, and 1 hours (Fu et al., 2017).

Table 5. Interaction effect of LED light regimes and rose cultivar on some flower characteristics.

| Cultivar | Light (LED day/night) Lux | Bud number | Mean harvest (weekly) | Cut flower height (cm) | Bud length (mm) | Bud diameter (mm) | Number of leaves | Stem diameter (mm) |
|----------------|---------------------------|------------|-----------------------|------------------------|-----------------|-------------------|------------------|--------------------|
| Allstar | Control | 4.25c | 1.12ab | 41.97e | 44.90e | 32.16e | 7.08c | 5.11f |
| | Night 2000 | 5.42ab | 1.01bc | 50.85d | 48.10d | 38.00bc | 8.83b | 6.13c |
| | Night 4000 | 5.41ab | 0.92c | 46.62de | 50.80cd | 38.63bc | 9.19b | 5.913d |
| | Day 2000 | 5.81a | 0.94c | 55.12cd | 52.03c | 34.70cd | 10.38ab | 6.75bc |
| | Day 4000 | 5.42ab | 1.11ab | 51.41d | 48.45d | 37.16bc | 9.73ab | 6.32c |
| | Night Day 2000 | 5.66ab | 0.92c | 58.42bcd | 51.29c | 35.16bcd | 9.43b | 6.68bc |
| | Night Day 4000 | 5.30bc | 0.93c | 55.19cd | 53.83c | 36.33bc | 9.50b | 6.27c |
| | Dark 2000 | 3.16d | 1.29a | 60.71ab | 44.86e | 34.03cd | 8.63b | 5.62e |
| | Dark 4000 | 3.08d | 1.03bc | 53.69cd | 42.23e | 32.06e | 8.04bc | 4.96g |
| | Dolcevita | Control | 4.47c | 0.68d | 47.21de | 52.20c | 34.91cd | 9.75ab |
| Night 2000 | | 5.21bc | 1.04bc | 66.28a | 56.66b | 39.63bc | 12.40a | 7.06ab |
| Night 4000 | | 6.36a | 1.03bc | 56.56cd | 58.93a | 40.00ab | 11.90a | 7.05ab |
| Day 2000 | | 5.70ab | 1.13ab | 54.00cd | 57.23ab | 40.00ab | 10.71ab | 6.91ab |
| Day 4000 | | 5.90a | 1.44a | 58.63bcd | 55.70b | 41.16a | 10.83ab | 6.88ab |
| Night Day 2000 | | 5.48ab | 1.25ab | 59.47abc | 55.53b | 41.30a | 12.07a | 6.90ab |
| Night Day 4000 | | 5.51ab | 1.00bc | 59.22abc | 58.90a | 40.73ab | 12.03a | 7.62a |
| Dark 2000 | | 2.33e | 1.03bc | 63.38ab | 49.95d | 38.84bc | 7.83c | 5.12f |
| Dark 4000 | | 2.03e | 1.23ab | 51.76d | 43.80e | 33.70c | 8.39b | 5.05f |

In each column, the averages that have at least one letter in common do not have a significant difference at the 5% probability level based on the least significant difference (LSD) test.

Alsanius et al, (2017) reported that red LED light 80% + blue 20% with 16 hours of illumination and intensity of 7000 lux increased the plant height and stem diameter of common sunflower, which is consistent with the present study. The effect of red-blue LED light at a ratio of 1:1 with an intensity of 17,000 lux on tomato (*Solanum lycopersicum*) increased the diameter of the plant stem and the height and growth of the root (Li et al., 2017; Alsanius et al., 2017).

Number of leaves, stem diameter, flower quality, greenness index, and chlorophyll fluorescence

In the interaction effect of light treatment with cultivar, the highest number of leaves was observed in the Allstar cultivar in LED day 2000 lux treatment, which was 47% more than the control, and in the Dolcevida cultivar in LED night 2000 lux treatment, which was 27% more than the control (Table 5).

In the interaction effect of light treatment with the cultivar, the maximum stem diameter was observed in the Allstar cultivar in LED day 2000 lux treatment, which was 34% compared with control. In the Dolcevida cultivar, the maximum stem diameter was observed in LED night day 4000 lux, which is a 35% increase in the stem diameter compared to the control, (Table 5). The stem diameter index with the indicators of the number of buds, length of buds, length of buds, number of leaves, quality of flowers, fresh weight, dry weight, and carbohydrates, a positive correlation was observed at the level of 1%, and a positive correlation was observed with the index after harvest at the level of 5%.

In the interaction effect of light treatment with cultivar, the highest flower quality was observed in the Allstar cultivar in LED day 4000 lux, which was 24% compared with control. The highest flower quality was observed in the Dolcevida cultivar in night 400 lux intensity, which was a 22% increase in flower quality compared with the control (Table 6). The flower quality with the parameters of bud number, bud length, bud diameter, stem diameter, number of leaves, fresh weight, dry weight, carbohydrate, and total chlorophyll, it is significant correlation at 1% level and the vase life index is significant at 5% level.

In the interaction effect of light treatment with the cultivar, the highest greenness was observed in the Allstar cultivar in LED night day 2000 lux, which showed a 10% increase in greenness compared with the control (Table 6). The greenness index has a positive correlation with the number of buds at the level of 1%, and a significant positive correlation was observed with anthocyanin and after harvest at the level of 5%.

In the interaction effect of light treatment with cultivar, the highest chlorophyll fluorescence was observed in the Allstar cultivar in LED day 4000 lux intensity, which was 7% compared with control. The highest chlorophyll fluorescence index of 4000 lux was observed in the Dolcevida cultivar at daylight (Table 6). The chlorophyll fluorescence index with weekly harvest, chlorophyll a, and total chlorophyll had a positive and significant correlation, and a negative correlation with thrips was observed at the 1% level.

According to Schroeter-Zakrzewska and Pradita's (2021) experiment, leaf color plays a crucial role in the quality of ornamental plants. They showed that light color significantly affected the SPAD index value in Chrysanthemum plants. Specifically, exposure to white + blue light and white and blue lights resulted in an increase in the SPAD index value. However, red + blue light exposure resulted in the highest index of greening leaves (SPAD) value and the shortest cuttings with the longest roots.

Fresh weight, dry weight, leaf surface, thrips pest symptom

In the interaction effect of light treatment with cultivar, the highest fresh weight of cut flowers was observed in the Allstar cultivar in LED night 4000 lux treatment, which was 43% more than the control. In the Dolcevit cultivar, the heaviest fresh weight of cut flowers was observed in LED night day 4000 lux, which is a 33% increase compared with control (Table 6). A positive correlation was observed with the indices of the number of buds, bud length, stem diameter, dry weight, and carbohydrate at the level of 1% and with the indices of vase life, thrips pest at the level of 5%.

In the interaction effect of light treatment with cultivar, the highest dry weight of cut flowers was observed in the Allstar cultivar in LED night 4000 lux treatment, which was 39% compared with control. The highest dry weight of cut flowers was observed in the Dolcevit cultivar in LED night day 4000 lux intensity, which is a 33% increase compared with control (Table 6). A positive correlation of 1% was observed in dry weight with weekly harvest indices, bud length, bud diameter, stem diameter, number of leaves, fresh weight, and carbohydrate. This finding is consistent with the findings of Fan et al. (2013), who investigated the effect of different intensities of LED light (50 % blue and 50 % red) on tomato (*Solanum lycopersicum*). Schroeter-Zakrzewska and Pradita's (2021) study found that Chrysanthemum plants exposed to white + blue light had the highest fresh and dry weights, averaging about 65.2 and 38.1 g, respectively. In contrast, the plants exposed to red + blue light had the lowest fresh weight (46.9 g) and dry weight (26.3 g).

In the interaction effect of light treatment with the cultivar, the highest leaf surface was observed in the Allstar cultivar in LED night day 2000 lux, and in the Dolcevit cultivar, the highest leaf surface was observed in LED day 2000 lux intensity (Table 6). The leaf surface has no significant correlation with other indicators.

Table 6. Interaction effect of LED light regimes and rose cultivar on some flower characteristics.

| Cultivar | Light | Flower quality | Greenness (SPAD) | Chlorophyll fluorescence (Fv/Fm) | F.W. of cut flowers (g) | D. W. of cut flowers (g) | Leaf surface (mm ²) | Thrips pest | Chlorophyll a of leaves (mg/g FW) |
|-----------|----------------|----------------|------------------|----------------------------------|-------------------------|--------------------------|---------------------------------|-------------|-----------------------------------|
| Allstar | Control | 6.88bc | 52.75ab | 0.72c | 5.63e | 33.22bc | 6723.60d | 8.00a | 0.27cde |
| | Night 2000 | 7.76ab | 51.26ab | 0.76ab | 6.60c | 37.75bc | 10499.90bdc | 5.66b | 0.28cd |
| | Night 4000 | 7.77ab | 52.30ab | 0.75ab | 8.03ab | 46.21a | 8779.30cde | 5.33bc | 0.31bc |
| | Day 2000 | 8.53a | 54.23ab | 0.77a | 7.84ab | 43.73abc | 12148.80ab | 4.66de | 0.35b |
| | Day 4000 | 8.00a | 53.70ab | 0.77a | 7.47bc | 41.23abc | 12146.60ab | 5.00cd | 0.45a |
| | Night Day 2000 | 7.99ab | 58.10a | 0.77a | 7.56bc | 43.42abc | 13317.80a | 5.66b | 0.22e |
| | Night Day 4000 | 8.16a | 53.33ab | 0.77a | 7.61bc | 44.52abc | 11865.50abc | 5.00cd | 0.36b |
| | Dark 2000 | 7.61ab | 48.63bc | 0.74b | 3.80f | 22.92c | 9353.60cde | 1.66e | 0.23e |
| Dark 4000 | 6.13b | 51.22ab | 0.74b | 3.99f | 23.04c | 11416.30abc | 2.00e | 0.29cd | |
| Dolcevit | Control | 7.10c | 50.33ab | 0.75ab | 6.47c | 39.51bc | 9925.00cde | 8.00a | 0.26de |
| | Night 2000 | 8.48a | 50.50ab | 0.77a | 8.42b | 48.52a | 12345.70ab | 5.00cd | 0.23e |
| | Night 4000 | 8.70a | 49.36bc | 0.75ab | 7.84ab | 44.91abc | 12124.20ab | 5.66b | 0.31bc |
| | Day 2000 | 8.23a | 48.96bc | 0.77a | 8.10ab | 45.12abc | 13040.20a | 4.00de | 0.24e |
| | Day 4000 | 8.34a | 50.13ab | 0.77a | 7.60bc | 44.22abc | 11613.50abc | 5.33bc | 0.32bc |
| | Night Day 2000 | 8.13a | 48.53bc | 0.77a | 7.91ab | 44.87bac | 12225.40ab | 5.51b | 0.38ab |
| | Night Day 4000 | 8.35a | 49.43bc | 0.77a | 8/60a | 48.44a | 12227.80ab | 2.33e | 0.38ab |
| | Dark 2000 | 6.39bc | 47.26b | 0.75ab | 6.23c | 37.35bc | 10577.90bcd | 2.00e | 0.22e |
| Dark 4000 | 6.63bc | 39.90c | 0.74b | 5.70e | 31.49bc | 14024.00a | 8.00a | 0.30bc | |

In each column, the averages that have at least one letter in common do not have a significant difference at the 5% probability level based on the least significant difference (LSD) test. F.W.: Fresh weight; D.W.: Dry weight.

The number of leaves in the LED light regimes and light intensity both increased the number of leaves in the rose. This finding is consistent with Duong et al. (2000) that reported that using LED light 70% red + 30% blue with an intensity of 2600 lux in conjunction with sunlight on strawberry plants can increase the number of leaves, stem and root height, and dry weight compared to plants grown under fluorescent light. Naznin et al. (2019) found that in pepper, leaf number was significantly increased under 95% red light with 5% blue LED compared to 100% red LED.

The plant weight of mint (*Mentha sativa*) and basil (*Ocimum basilicum*) increased at LED light 70% red and 30% blue with 18 hours of lighting 4 times (Sabzalian et al. 2014). Tomatoes were grown under 1:1 blue and red LED light conditions, and the specific leaf surface area and total chlorophyll increased, resulting in better light absorption (Fan et al. 2013).

In the interaction effect of light treatment with cultivar, the highest thrips pest was observed in the control with an average of 8 out of 10 and the lowest thrips drop was observed in LED dark 2000 lux intensity with an average of 1.66. In the Delsovita cultivar, the highest thrips infestation was observed in control and the lowest thrips drop was observed in LED dark 2000 lux (Table 6). There was a positive correlation between the thrips pest index and the number of buds at the 1% level, and a negative correlation was observed with the number of flowers and chlorophyll fluorescence at the 1% level, and a significant correlation observed with the greenness index, fresh weight, and dry weight.

Johansen et al., (2018) evaluated the phototactic response of *F. occidentalis* to yellow and blue sticky traps with blue LED in colored and white *Alstroemeria* varieties. They found that catches in blue traps with LED were 3.4 and 4.0 times higher compared to blue traps without LED. In contrast, catches in yellow traps with LED increased by 4.5 times compared to yellow traps without LED, but they were only slightly higher than those observed in blue traps without light and lower than catches in blue traps with LED. This difference is likely due to the reflection of blue light in the blue sticky traps, which produced a higher stimulus, while the blue light is mostly absorbed in the yellow traps.

Physiological factors chlorophyll a, chlorophyll b, total chlorophyll, carotenoids, petal anthocyanin content

In the interaction effect of light treatment with cultivar, the highest chlorophyll a was observed in the Allstar cultivar in LED day 4000 lux, which was 66% compared with control. Delsovita cultivar in LED night day 2000 lux, which is a 46% increase compared with control (Table 6). The chlorophyll a with chlorophyll b, total chlorophyll, and carotenoid indices is significant at 1% level and with life after harvest at 5% level.

In the interaction effect of light treatment with cultivar, the highest chlorophyll b was observed in the Allstar cultivar in LED day 4000 lux, which was 60% compared with the control. The most chlorophyll b was observed in the Delsovita cultivar in LED night day 4000 lux, which is a 42% increase compared to the control (Table 7). The chlorophyll b with total chlorophyll, carotenoids, and after harvest, a positive correlation was observed at the level of 1%, and with the number of leaves and water content, a positive correlation was observed at the level of 5%.

In the interaction effect of light treatment with the cultivar, the highest total chlorophyll was observed in the Allstar cultivar in LED day 4000 lux, which was 63% compared with control. The highest total chlorophyll was observed in the Dolcevita cultivar in LED night day 4000 lux, which is a 44% increase compared to the control (Table 7). The total chlorophyll with carotenoid indicators and after harvesting, a positive correlation was observed at the

level of 1%, and with the content of flower water, carbohydrates, and total chlorophyll, a positive correlation was observed at the level of 5%.

In the interaction effect of light treatment with cultivar, the most carotenoid was observed in the Allstar cultivar in LED day 4000 lux, which was 64% compared with control. The most carotenoids were observed in the Dolcevita cultivar in LED night day 2000 lux, which is a 62% increase compared to the control (Table 7). A positive correlation was observed between carotenoids and total chlorophyll indices after harvest, at the level of 1%, and a positive correlation was observed with the content of flower water content and carbohydrates at the level of 5%.

In the interaction effect of light treatment with cultivar, the highest anthocyanin content was observed in the Allstar cultivar in LED day 2000 lux, which was 41% higher than the control. The highest anthocyanin content was observed in the Delsovita cultivar in LED night 2000 lux, which was a 113% increase compared with control (Table 7). The anthocyanin with greenness index at the level of 1% and with the vase life index, carbohydrate, and flower water content, a positive correlation was observed at the level of 5%.

Chlorophylls try to adapt to the environment in different environmental conditions so that the chlorophyll content changes to improve maximum photon absorption in different environmental conditions (Samuoliene et al., 2011). When a plant is exposed to blue light in addition to other wavelengths, its chlorophyll content increases (Hernandez, 2013). According to Wu et al. (2007), pigments, particularly chlorophyll and carotenoid, decreases in the dark. The duration of light exposure can also affect the chlorophyll content. However, it has been determined that red light causes carotenoid and anthocyanin accumulation while blue light causes chlorophyll accumulation. Anthocyanin biosynthesis is typically linked to blue light via cryptochromes. However, active phytochrome is required for the regulation of anthocyanin synthesis via cryptochromes, and this requirement can be related to the presence of phytochrome a or phytochrome b (Ahmad & Cashmore, 1997).

According to Zhang et al. (2020), increasing the exposure time to light results in an increase in chlorophyll and carotenoids in microgreens, which is consistent with the current study. Fan et al. (2013) found that exposing cabbage to a red-blue LED light with a 6:1 ratio of 8000 lux without sunlight for 12 hours on and 12 hours off increased carotenoids by about 70% when compared to mono spectral lights.

The effect of LED and cultivar on the vase life of two rose cultivars

Soluble carbohydrate, vase life parameters, and the speed of flower bud opening

In the interaction effect of light treatment with cultivar, the highest carbohydrate was observed in the Allstar cultivar in LED day 4000 lux, which was 31% compared with control. In the Dolcevita cultivar, the highest carbohydrates were observed in LED night day 4000 lux, which is a 142% increase compared with control (Table 7). The carbohydrate content has a positive correlation with the number of buds, bud length, bud diameter, stem diameter, number of leaves, flower quality, fresh weight, and dry weight at the level of 1%, and with chlorophyll and water content, there is a positive and significant correlation at the level of 5% was observed.

Growing tomatoes under 70% red + 30% blue light without sunlight with a light intensity of 15,000 lux was shown by Li et al. (2017) to increase total carbohydrates and starch, as well as sucrose accumulation. Hao et al. (2016) found that red-blue supplementary LED light at a ratio of 1:1 with an intensity of 4700 lux increased the number of flower buds as well as the amount of carbohydrates, which is consistent with the current study.

In the interaction effect of light treatment with cultivar, the longest vase life was observed in the Allstar cultivar in LED night day 4000 lux, which is 44% compared with control. In the

Dolcevita cultivar, the longest vase life was observed in LED day 4000 lux, which is a 133% increase compared with control (Table 7). The vase life index has a positive correlation with the number of buds, chlorophyll b, total chlorophyll, carotenoids, and anthocyanins at the 1% level, as well as with stem diameter, flower quality, greenness, fresh weight, dry weight, flower water content, chlorophyll A positive correlation was observed at the 5% level.

Horticultural products have a limited shelf life after harvesting, which is due to factors such as weight loss, aging, loss of strength, softening of tissue corruption, and so on. Supplemental lighting during plant growth can increase vase life by assisting in the storage of sugars and carbohydrates (Hasperue et al., 2016). The use of LED light has a significant impact on the life of horticultural products after harvest. In its most basic form, artificial LED lights can help with more photosynthesis assimilate and more food storage (Dayani et al., 2018). According to Samuoliene et al. (2012), in a study of the effect of red LED light on the changes in photochemical content in lettuce leaves, it was discovered that exposing lettuce plants to LED light during cultivation increased the production of ascorbic acid, which has antioxidant properties and reduces the activity of radicals. In the same study, the carbohydrate content and antioxidant capacity of the lettuce plant increased under the LED light treatment, which had a positive correlation with the increase in vase life after harvesting the plant (Samuoliene et al., 2012).

A study found that using a 1000 lux LED supplemental light increased the vase life of cabbage after harvesting and delayed the yellowing of its inflorescences (Hasperue et al., 2016). It was able to increase the vase life of the plant after harvesting by 30% more than the Allstar control and 23% more than the Dolsovit control in the vase life light intensity index of 4000 lux, which is consistent with the research of Pettersen et al. (2007) who reported that increasing the exposure time to the plant increased the longevity of flowers on potted roses.

Table 7. Interaction effect of LED light regimes and rose cultivar on some flower characteristics.

| Cultivar | Light | Chlorophyll b of leaves (mg/g FW) | Total Chlorophyll of leaves (mg/g FW) | Carotenoids of leaves (mg/g FW) | Anthocyanin of petal (μmol/g) | Carbohydrate of leaves (mg/g) | Vase life (day) | Speed of flower bud opening (mm/day) | Flower water content (%) |
|----------------|----------------|-----------------------------------|---------------------------------------|---------------------------------|-------------------------------|-------------------------------|-----------------|--------------------------------------|--------------------------|
| Allstar | Control | 0.20e | 0.73bc | 0.17cde | 227.68bcd | 124.05de | 9.00de | 2.021c | 81.21b |
| | Night 2000 | 0.21de | 0.77bc | 0.18cde | 283.84ab | 148.00d | 11.00c | 2.22bc | 82.90ab |
| | Night 4000 | 0.25ab | 0.87b | 0.20bc | 310.81a | 160.50a | 11.66c | 2.29bc | 82.73ab |
| | Day 2000 | 0.24bc | 0.93b | 0.23b | 322.73a | 152.69d | 12.33b | 2.73a | 83.24a |
| | Day 4000 | 0.32a | 1.19a | 0.28a | 307.58a | 163.23cd | 12.33b | 2.82a | 82.65ab |
| | Night Day 2000 | 0.19efg | 0.73bc | 0.15e | 258.29abc | 159.25d | 11.66c | 2.82a | 83.11a |
| | Night Day 4000 | 0.29a | 0.99b | 0.23b | 258.89abc | 157.30d | 13.00a | 2.76a | 82.45ab |
| | Dark 2000 | 0.20e | 0.65d | 0.15e | 163.43de | 125.706de | 10.00d | 2.16c | 81.84b |
| | Dark 4000 | 0.22cd | 0.78bc | 0.19cd | 194.75cd | 129.75de | 9.66de | 2.73a | 82.00ab |
| | Dolcevita | Control | 0.19efg | 0.70cd | 0.16e | 13.99g | 74.14f | 5.00e | 1.79f |
| Night 2000 | | 0.21de | 0.66d | 0.14e | 29.83g | 155.40d | 10.00d | 2.61abc | 83.38a |
| Night 4000 | | 0.22cd | 0.83b | 0.19cd | 21.21g | 168.18abc | 11.33c | 2.27bc | 83.17a |
| Day 2000 | | 0.17g | 0.64d | 0.15e | 19.80g | 163.37cd | 11.66c | 2.87a | 83.40a |
| Day 4000 | | 0.24bc | 0.87b | 0.21bc | 24.34g | 176.30ab | 11.66c | 2.64abc | 83.46a |
| Night Day 2000 | | 0.26ab | 1.01a | 0.26a | 17.37g | 167.19bc | 10.66d | 2.94a | 83.11a |
| Night Day 4000 | | 0.27a | 1.01a | 0.25a | 22.32g | 179.84a | 8.33de | 2.59bcd | 83.49a |
| Dark 2000 | | 0.19efg | 0.63d | 0.13e | 11.52g | 138.91d | 10.00d | 3.00a | 82.60ab |
| Dark 4000 | | 0.21de | 0.79bc | 0.18cde | 12.13g | 147.63d | 11.33c | 2.05ef | 82.30ab |

In each column, the averages that have at least one letter in common do not have a significant difference at the 5% probability level based on the least significant difference (LSD) test.

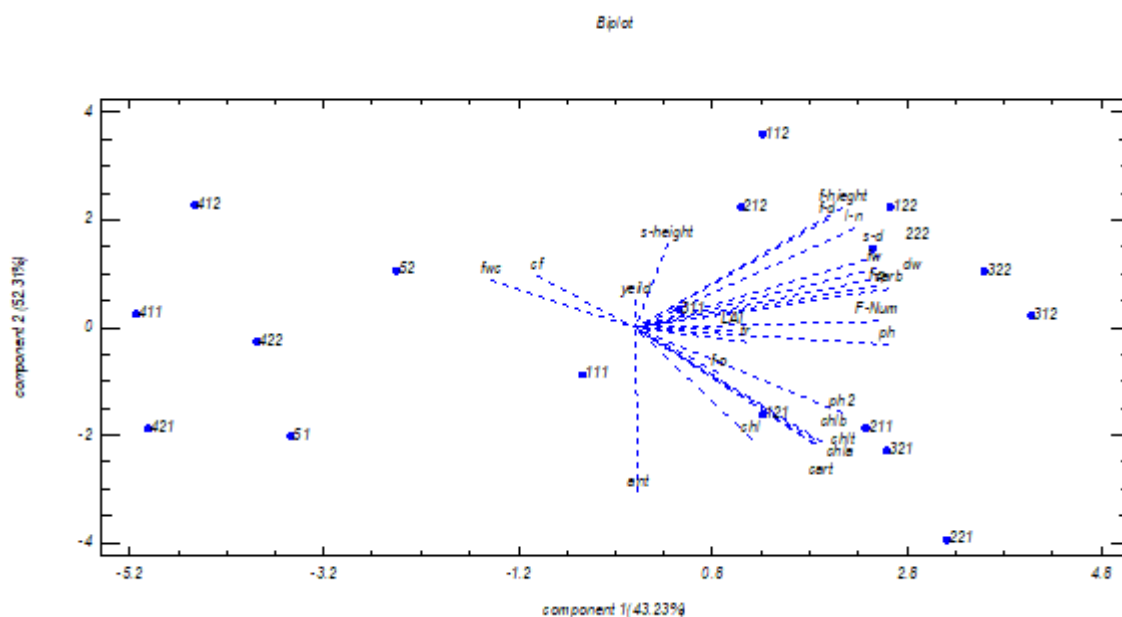


Fig. 3. The PCA analysis of the effect of LED on some characteristics of Rose.

Bud number (F-Nun), Mean harvest weekly (yield), Cut flower height (S-height), Bud diameter (F-d), Stem diameter (S-d), Number of leaves (l-n), Flower quality (f-q), Greenness (Chl), Chlorophyll fluorescence (cf), Fresh weight (fw), Dry weight (dw), Flower water content (fwc), Vase life 1 day (ph), Vase life 2 day (ph2), Chlorophyll a (Chla), Chlorophyll b (Chlb), Total chlorophyll (Chlt), Carotenoid (Cart), Anthocyanin (ant), Leaf surface (LA1), Thrips pest (tr), Speed of flower bud opening (f-o), Carbohydrate of leaves (Carb). N2 Allstar (111), N4 Allstar (121), D2 Allstar (211), D4 Allstar (221), DN2 Allstar (311), DN4 Allstar (321), T2 Allstar (411), T4 Allstar (421), Control1 Allstar (51), N2 Dolcevita (112), N4 Dolcevita (122), D2 Dolcevita (212), D4 Dolcevita (222), ND2 Dolcevita (312), ND4 Dolcevita (322), T2 Dolcevita (412), T4 Dolcevita (422), Control 11 Dolcevita (52).

In the interaction effect of light treatment with the cultivar, the Allstar cultivar had the fastest flower opening in LED day 4000 lux, which was 40% faster than the control (Table 7).

Flower water content was highest in LED day 2000 lux in Allstar and in night day 4000 lux in Dolcevita cultivar (Table 7).

According to PCA the best light intensity for increasing growth parameters and pigment content with 4000 lux in both cultivars and the best LED regime was 24 hours on (with sunlight), 24 hours light (without sunlight) in both cultivars improve the majority of flowers characteristics (Fig. 3).

CONCLUSION

Conclusively, chlorophyll parameters such as chlorophyll a, b, total, carotenoid, chlorophyll fluorescence and also the average number of flower harvests per week improved in LED day 4000 lux. The light intensity of 4000 lux had a better effect on the vase life of roses in both cultivars. Allstar showed the longest vase life in the treatment of LED night day 4000 lux, and Dolcevita showed the longest vase life in LED day 4000 lux treatment. In general, according to the information obtained in this research, it is recommended to use 4000 lux LED supplemental light during the day to increase the quantity and quality of roses in low light seasons (autumn and winter).

Conflict of interest

To the best of our knowledge, the named authors have no conflict of interest, financial or otherwise.

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REFERENCES

- Aalifar, M., Aliniaiefard, S., Arab, M., Zare Mehrjerdi, M., Dianati Daylami, S., Serek, M., Woltering, M., & Li, T. (2020). Blue light improves vase life of carnation cut flowers through its effect on the antioxidant defense system. *Frontiers in Plant Science*, *11*, 511. <https://doi.org/10.3389/fpls.2020.00511>.
- Ahmad, M., & Cashmore, A.R. (1997). The blue-light receptor cryptochrome 1 shows functional dependence on phytochrome A or phytochrome B in *Arabidopsis thaliana*. *The Plant Journal*, *11*, 421-427. <https://doi.org/10.1046/j.1365-313X.1997.11030421.x>.
- Alaey, M., Babalar, M., Naderi, R., & Kafi, M. (2011). Effect of pre-and postharvest salicylic acid treatment on physio-chemical attributes in relation to vase-life of rose cut flowers. *Postharvest Biology and Technology*, *61*(1), 91-94. <https://doi.org/10.1016/j.postharvbio.2011.02.002>.
- Alsanius, B.W., Bergstrand, K.J., Hartmann, R., Gharaie, S., Wohanka, W., Dorais, M., & Rosberg, A.K. (2017). Ornamental flowers in new light: artificial lighting shapes the microbial phyllosphere community structure of greenhouse grown sunflowers (*Helianthus annuus* L.). *Scientia Horticulturae*, *216*, 234-247. <https://doi.org/10.1016/j.scienta.2017.01.022>.
- Arnon, D.I. (1949). Copper enzymes in isolated chloroplasts. polyphenoloxidase in *Beta vulgaris*. *Plant Physiology*, *24*(1), 1-15.
- Carotti, L., Potente, G., Pennisi, G., Ruiz, K.B., Biondi, S., Crepaldi, A., Orsini, F., Gianquinto, G., & Antognoni, F. (2021). Pulsed LED light: exploring the balance between energy use and nutraceutical properties in indoor-grown lettuce. *Agronomy*, *11*, 1106. <https://doi.org/10.3390/agronomy11061106>.
- Cioć, M., Dziurka, M., & Pawłowska, B. (2022). Changes in endogenous phytohormones of gerbera jamesonii axillary shoots multiplied under different light emitting diodes light quality. *Molecules*, *27*(6), 1804. <https://doi.org/10.3390/molecules27061804>.
- Dayani, S., Heydarzadeh, P., & Sabzalian, M.R. (2018). *Efficiency of light-emitting diodes for future photosynthesis*. In Handbook of photosynthesis (pp. 761-783). CRC press.
- Dezhabad, F., & Haghghi, M. (2020). Bottom-cold stress was less harmful than cold-air stress on tomato seedling production treated with boric acid. *Acta Physiologiae Plantarum*, *42*(4), 44. <https://doi.org/10.1007/s11738-020-3035-2>.
- Duong, T.N., Takamura, T., Watanabe, H., & Tanaka, M. (2000). *Light emitting diodes (LEDs) as a radiation source for micropropagation of strawberry*. Transplant Production in the 21st Century. Springer. Dordrecht. (pp. 114-118).
- Fan, X.X., Xu, Z.H., Liu, X.Y., Tang, C.M.L., Wang, W., & Han, X.L. (2013). Effects of light intensity on the growth and leaf development of young tomato plants grown under a combination of red and blue light. *Scientia Horticulturae*, *153*, 50-55. <https://doi.org/10.1016/j.scienta.2013.01.017>.
- Fu, Y., Li, H., Yu, J., Liu, H., Cao, Z., Manukovsky, N.S., & Liu, H. (2017). Interaction effects of light intensity and nitrogen concentration on growth, photosynthetic characteristics and quality of lettuce (*Lactuca sativa* L. Var. Youmaicai). *Scientia Horticulturae*, *214*, 51-57. <https://doi.org/10.1016/j.scienta.2016.11.020>.

- Gao, D., Ji, X., Yuan, Q., Pei, W., Zhang, X., Li, F., Han, Q., & Zhang, S. (2023). Effects of total daily light integral from blue and broad-band red LEDs on flowering of saffron (*Crocus sativus* L.). *Scientific Reports*, *13*, 7175. <https://doi.org/10.1038/s41598-023-34424-0>.
- Hao, X., Little, C., Zheng, J.M., & Cao, R. (2016). Far-red LEDs improve fruit production in greenhouse tomato grown under high-pressure sodium lighting. *Acta Horticulture*, *134*, 95–102. <https://doi.org/10.17660/ActaHortic.2016.1134.13>.
- Hasperue, J.H., Rodoni, L.M., Guardianelli, L.M., Chaves, A.R., & Martínez, G.A. (2016). Use of LED light for brussels sprouts postharvest conservation. *Scientia Horticulturae*, *213*, 281–286. <https://doi.org/10.1016/j.scienta.2016.11.004>.
- Hedge, J.E., & Hofreiter, B.T. (1962). *Carbohydrate chemistry 17*. In Whistler, R.L. and Be Miller, J. N., Eds., Academic Press, New York.
- Heo, J.W., Lee, C.W., Murthy, H.N., & Paek, K.Y. (2003). Influence of light quality and photoperiod on flowering of *Cyclamen persicum* Mill. cv. 'Dixie White'. *Plant Growth Regulation*, *40*, 7-10. <https://doi.org/10.1023/A:1023096909497>.
- Hernandez, R. (2013). *Growth and development of greenhouse vegetables Seedlings under supplemental LED lighting*. The University of Arizona. Arizona.
- Iacona, C., & Muleo, R. (2010). Light quality affects in vitro adventitious rooting and ex vitro performance of cherry rootstock Colt. *Scientia Horticulturae*, *125*(4), 630-636. <https://doi.org/10.1016/j.scienta.2010.05.018>.
- Johansen, N.S., Torp, T., & Solhaug, K.A. (2018). Phototactic Response of *Frankliniella occidentalis* to sticky traps with blue light emitting diodes in herb and alstroemeria greenhouses. *Crop Protection*, *114*, 120–128.
- Kurepin, L.V., Walton, L.J., Yeung, E.C., Chinnappa, C.C., & Reid, D.M. (2010). The interaction of light irradiance with ethylene in regulating growth of *Helianthus annuus* shoot tissues. *Plant Growth Regulation*, *62*, 43-50. <https://doi.org/10.1007/s10725-010-9483-8>.
- Lee, C.G., & Palsson, B.Ø. (1994). High-density algal photobioreactors using light-emitting diodes. *Biotechnology & Bioengineering*, *44*(10), 1161-1167. <https://doi.org/10.1002/bit.260441002>.
- Li, Y., Xin, G., Wei, M., Shi, Q., Yang, F., & Wang, X. (2017). Carbohydrate accumulation and sucrose metabolism responses in tomato seedling leaves when subjected to different light qualities. *Scientia Horticulturae*, *225*, 490-497. <https://doi.org/10.1016/j.scienta.2017.07.053>.
- Livadariu, O., Maximilian, C., Rahmanifar, B., & Cornea, C.P. (2023). LED technology applied to plant development for promoting the accumulation of bioactive compounds: a review. *Plants*, *12*(5), 1075. <https://doi.org/10.3390/plants12051075>.
- Maxwell, K., & Johnson, G.N. (2000). Chlorophyll fluorescence a practical guide. *Journal of Experimental Botany*, *51*, 659-668. <https://doi.org/10.1093/jexbot/51.345.659>.
- Naznin, M.T., Lefsrud, M., Gravel, V., & Azad, M.O.K. (2019). Blue light added with red LEDs enhance growth characteristics, pigments content, and antioxidant capacity in lettuce, spinach, kale, basil, and sweet pepper in a controlled environment. *Plants*, *8*(4), 93.
- Nikbakht, A., & Ashrafi, N. (2019). *Cut flowers: practical and scientific growing*. Isfahan. Iran: Isfahan University of Technology. (In Persian).
- Park, Y.G., & Jeong, B.R. (2020). How supplementary or night-interrupting low-intensity blue light affects the flower induction in chrysanthemum, a qualitative short-day plant. *Plants*, *9*(12), 1694.
- Pérez-Grajales, M., Martínez-Damián, M., Cruz Álvarez, O., Potrero-Andrade, S., Peña Lomeli, A., González-Hernández, V., & Villegas-Monter, A. (2019). Content of capsaicinoids and physicochemical characteristics of Manzano hot pepper grown in greenhouse. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, *47*(1), 119-127. <https://doi.org/10.15835/nbha47111241>.
- Pettersen, R.I., Moe, E., & GIslerod, H.R. (2007). Growth of pot roses and postharvest rate of water loss as affected by air humidity and temperature variations during growth under continuous light. *Scientia Horticulturae*, *114*, 207–213. <https://doi.org/10.1016/j.scienta.2007.06.009>.
- Rapisarda, P., Fanella, F., & Maccarone, E. (2000). Reliability of analytical methods for determining anthocyanins in blood orange juices. *Journal of Agricultural and Food Chemistry*, *48*(6), 2249-2252. <https://doi.org/10.1021/jf991157h>.

- Rasouli, O., Ahmadi, N., Behmanesh, M., & Nergi, M.D. (2015). Effects of BA and TDZ on postharvest quality and expression of laccase and aquaporin genes in cut rose 'Sparkle'. *South African Journal of Botany*, 99, 75-79. <https://doi.org/10.1016/j.sajb.2015.03.191>.
- Sabzalian, M.R., Heydarizadeh, P., Zahedi, M., Boroomand, A., Agharokh, M., Sahba, M.R., & Schoefs, B. (2014). High performance of vegetables, flowers, and medicinal plants in a red-blue LED incubator for indoor plant production. *Agronomy for Sustainable Development*, 34, 879–886. <https://doi.org/10.1007/s00425-004-1418-z>.
- Sakurako, H., Shota, Y., Haruki, K., Saashia, F., Shigekazu, K., Ken-Ichiro, S., & Atsushi, T. (2021). A BLUS1 kinase signal and a decrease in intercellular CO₂ concentration are necessary for stomatal opening in response to blue light. *Plant Cell*, 33, 1813–1827.
- Samuoliene, G., Brazaityte, A., Sirtautas, R., Novickovas, A., & Duchovskis, P. (2011). Supplementary red-LED lighting affects phytochemicals and nitrate of baby leaf lettuce. *Journal of Food, Agriculture and Environment*, 9, 271-274.
- Samuoliene, G., Sirtautas, R., Brazaityte, A., Virsile, A., & Duchovskis, P. (2012). Supplementary red-LED lighting and the changes in phytochemical content of two baby leaf lettuce varieties during three seasons. *Journal of Food, Agriculture and Environment*, 10, 701-706.
- Schroeter-Zakrzewska, A., & Pradita, F.A. (2021). Effect of colour of light on rooting cuttings and subsequent growth of chrysanthemum (*Chrysanthemum × grandiflorum* Ramat./Kitam.). *Agriculture*, 11(7), 671. <https://doi.org/10.3390/agriculture11070671>.
- Shi, L., He, S., Wang, Z., & Kim, W.S. (2021). Influence of nocturnal supplemental lighting and different irrigation regimes on vase life and vase performance of the hybrid rose 'Charming Black'. *Horticultural Science and Technology*, 39(1), 23-36.
- Song, Y., Shang, W., Ma, D., Wang, Z., He, S., Shi, L., Shen, Y., He, D., Wang, E., & Wang, X. (2022). Effect on the growth and photosynthetic characteristics of *Anthurium andreanum* ('Pink Champion', 'Alabama') under hydroponic culture by different LED light Spectra. *Horticulturae*, 8(5), 389. <https://doi.org/10.3390/horticulturae8050389>.
- Wu, M.C., Hou, C.Y., Jiang, C.M.Y., Wang, T.C., Wang, Y.H., Chen, H.H., & Chang, M. (2007). A novel approach of LED light radiation improves the antioxidant activity of pea seedlings. *Food Chemistry*, 101, 1753-1758. <https://doi.org/10.1016/j.foodchem.2006.02.010>.
- Zhang, X., Bian, Z., Yuan, X., Chen, X., & Lu, C. (2020). A review on the effects of light-emitting diode (LED) light on the nutrients of sprouts and microgreens. *Trends in Food Science & Technology*, 99, 203-216.

